

Universität
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Traditio et Innovatio



Universitätsmedizin
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HAUPT- G A N G

What is wrong with my flow cytometry data?

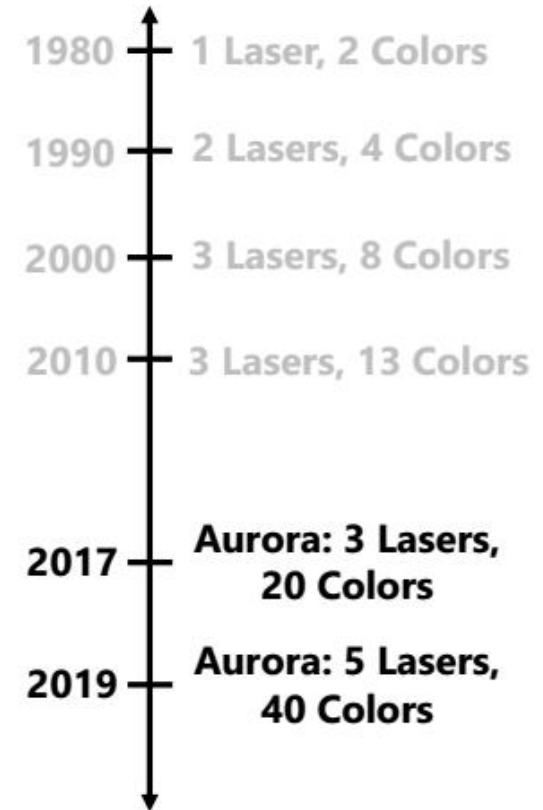
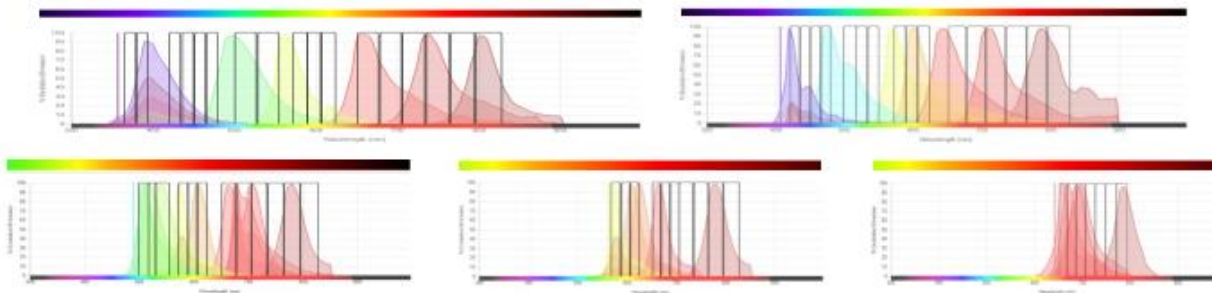
Hints, tricks and pitfalls

Webinar 7
01.08.2023



Golden rules of panel design

The more...the better (the more complicated)



source: Jesus Gil Pulido - Cytex Training Rostock 2020

What do you want to look at...

1. Which type of cells you want to look at?
 1. Mouse/human/...
 2. Origin (blood, tissue,...)
 3. Antigens
 4. Gating strategy
2. What is my sample's nature?
 1. Live or fixed
 2. Rare cells
 3. Small amount
3. What is the configuration of my analyzer?
 1. Number and type of lasers
 2. Number and bandwidth of filters



3 types of antigens

If the level of expression of your target protein is unknown then consider antigen as tertiary!

Primary

High density
on or off expression

CD3
CD4
CD8

Secondary

Intermediate density
continuous expression

CD45RA
CCR7
CD127
HLA-DR

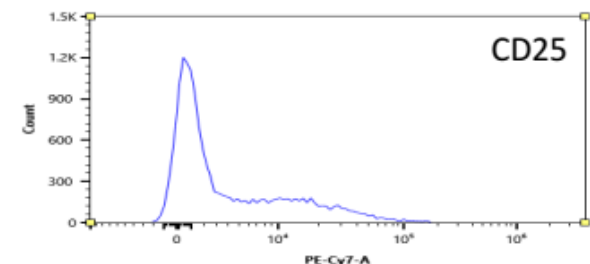
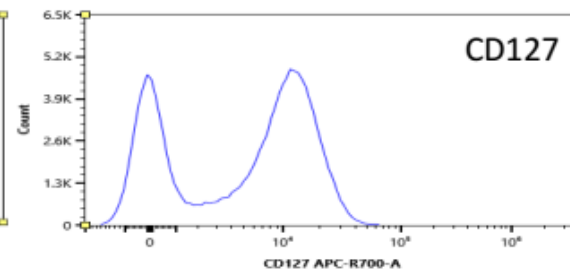
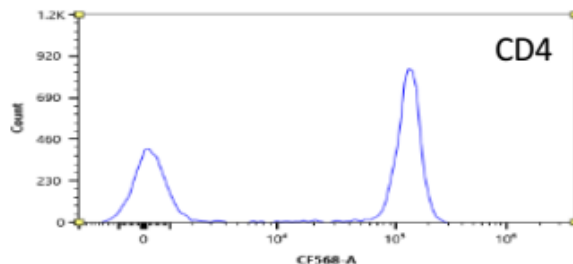
Tertiary

Low density
unknown expression

PD-1
CD25
TCR γ/δ

Fluorochrome Brightness

Level of Antigen Expression



Mahnke, Y. and Roederer, M. Clin Lab Med. 2007 September ; 27(3): 469

direction of choosing fluorophors (searching for products)

source: Jesus Gil Pulido - Cytek Training Rostock 2020

REMINDER: Check fluorophore brightness

Laser	Fluorochrome			
	Very Bright	Bright	Moderate	Dim
	Ultraviolet (355 nm)	BD Horizon™ BUV563 BD Horizon™ BUV615 BD Horizon™ BUV661 BD Horizon™ BUV737	BD Horizon™ BUV395 BD Horizon™ BUV496	BD Horizon™ BUV805
	Violet (405 nm)	BD Horizon™ BV421 BD Horizon™ BV650 BD Horizon™ BV711	BD Horizon™ BV480 BD Horizon™ BV605 BD Horizon™ BV786	BD Horizon™ V450 BD Horizon™ V500
	Blue (488 nm)	BD Horizon™ BB515 BD Horizon™ BB700 BD Horizon™ PE-CF594 PE-Cy™5	PE PE-Cy™7 FITC Alexa Fluor® 488 PerCP-Cy™5.5	PerCP
	Yellow/Green (561 nm)	PE BD Horizon™ PE-CF594 PE-Cy™5 PE-Cy™7		
	Red (640 nm)		APC Alexa Fluor® 647 BD Horizon™ APC-R700 BD Horizon™ R718	Alexa Fluor® 700 APC-H7 APC-Cy™7

https://www.bdl.co.jp/biosciences/support/pdf/23-16181_Post_Card_BD_Life_Sciences_Relative_Fluorochrome_Brightness.pdf

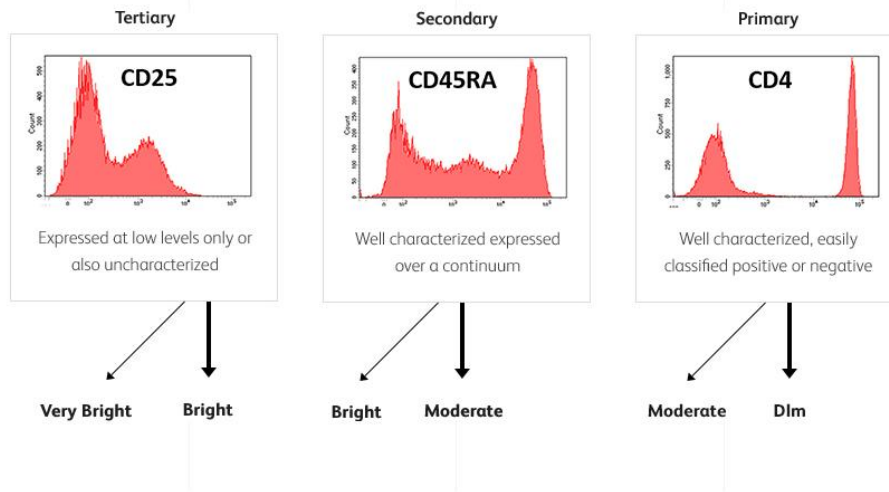
		APC-Cy™7	Em (nm) Max	Filter Used	Brightness
https://www.biollegend.com/en-us/brightness-index	PE	565	575	585/20	5
	PE/Cyanine5	565	670	660/20	5
	PE/Dazzle™ 594	565	610	610/20	5
	PE/Fire™ 700*	565	695	670/30	5
	Brilliant Violet 421™	405	421	450/50	5
	Brilliant Violet 711™	405	711	710/50	5
	Brilliant Violet 605™	405	603	610/20	4
	Brilliant Violet 650™	405	645	660/20	4

<https://www.biollegend.com/en-us/brightness-index>

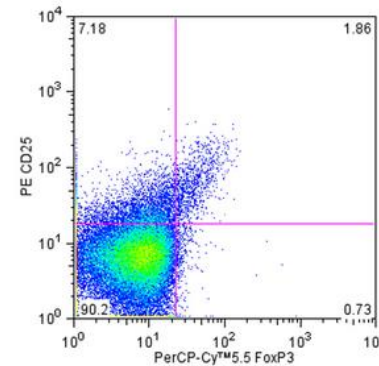
REMINDER: check spillover-spread

Match antigen expression with fluorochrome brightness

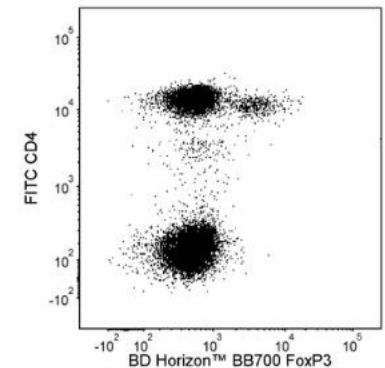
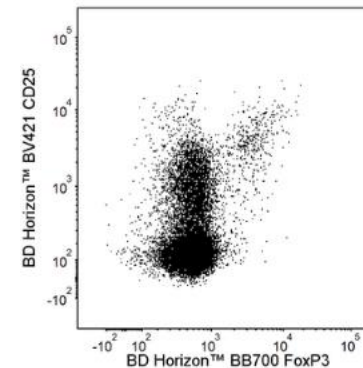
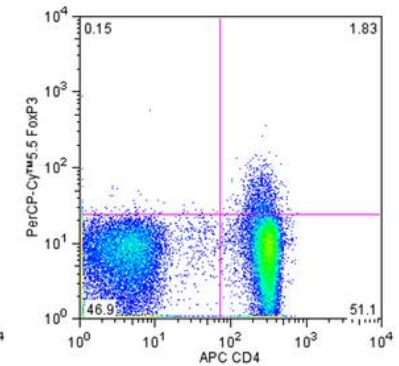
Match tertiary antigens with bright fluorochromes and primary antigens with dim fluorochromes. In the following table, you can find Fluorochrome ranking on brightness which can be used to help you in panel design.



CD25:PE and FoxP3:PerCP/Cy5.5 co-expression / overlap → bad resolution



CD4:APC to bright → much spread → bad resolution of PerCP/Cy5.5



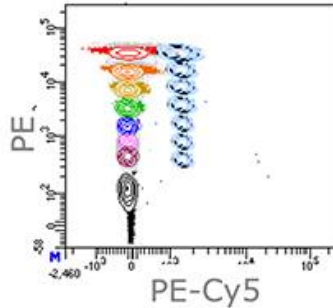
<https://eu.bd.com/panel-design/en/flow-cytometry-fluochrome-brightness-spillover>

<https://wwwbdbiosciences.com/en-de/products/reagents/flow-cytometry-reagents/research-reagents/single-color-antibodies-ruo/percp-cy-5-5-mouse-anti-human-foxp3.561493>

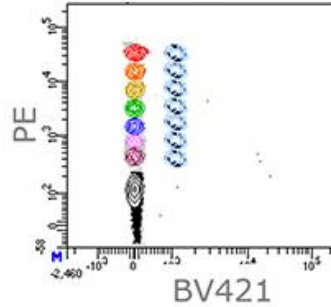
<https://wwwbdbiosciences.com/en-de/products/reagents/flow-cytometry-reagents/research-reagents/single-color-antibodies-ruo/bb700-mouse-anti-human-foxp3.566527>

REMINDER: check spillover-spread

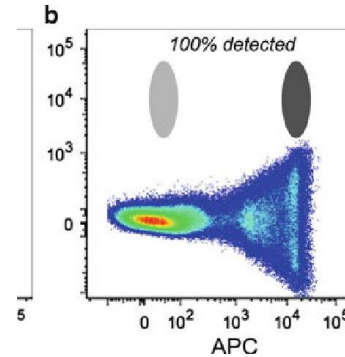
Comp. 9.15%



0.0%



BD Comp Beads stained with increasing amount of PE antibody to get increasing MFIs of pos. populations.



Less spillover increases overall quality of your panel.

- (very) bright dyes have increased emitted photon amount
- increase of spillover spread
- can reduce resolution especially for co-expressed markers (double positive)

<https://eu.bd.com/panel-design/en/flow-cytometry-fluochrome-brightness-spillover>

Golden Rules of Panel design – step by step

1. Which type of cells you want to look at?
 - literature search: already described panels → OMIPs
 - OMIPs = Optimized Multicolor Immunofluorescence Panel published e.g. in Cytometry Part A
 - antibodies you already have in the lab OR without alternative
 2. Search for antibodies for tertiary antigens (intranuclear, cytoplasmatic proteins)
 - most important targets
 - use the brightest dyes (PE, BV421, APC, PE-Cy7; ...)
 - spread the antibodies over all lasers to reduce needed compensation
- Fill your panel

Golden Rules of Panel design – step by step

3. Combine and add highly and moderate markers with dim dyes

- CD3-FITC, CD45-PerCP, ...

➤ Fill your panel

3. Include live-dead stain

- fixable (Zombie, FVS...)
- non-fixable (PI, 7-AAD, DAPI)

➤ Fill your panel

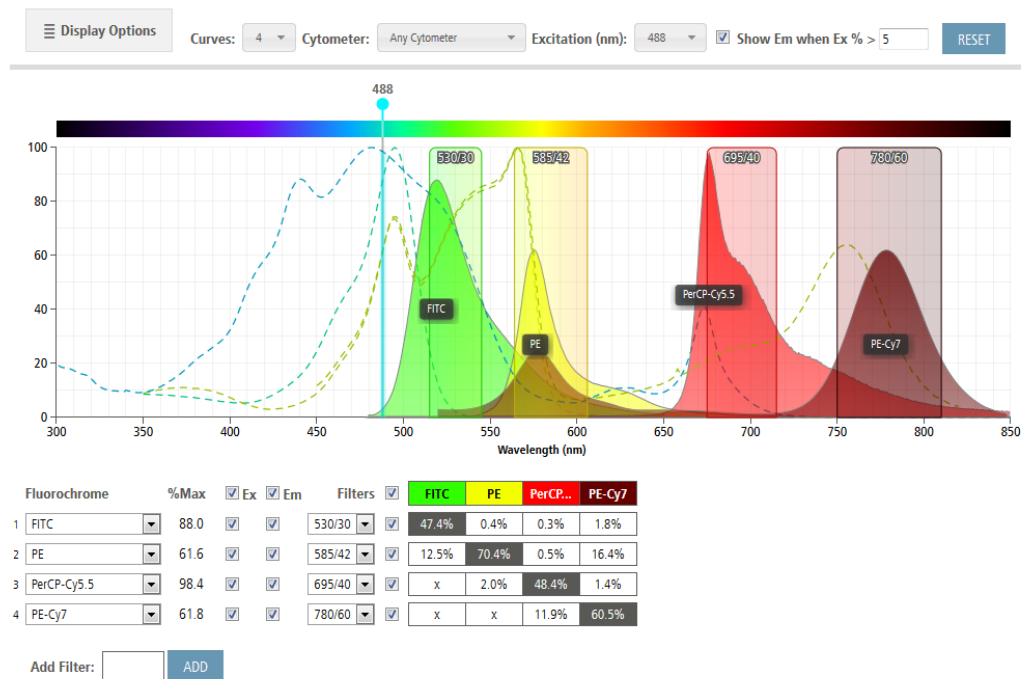
3. Check your panel:

- put markers on highly autofluorescent cells in red laser module
- live-dead stain and ubiquitous markers at the edges of the spectrum (less compensation)
- compare antibody clones (resolution)
- use better (tandem-) dyes with less spread (e.g. PerCP → PerCP-Cy5.5 or BB700 or PerCP-Vio700)
- check for fixability of dyes (use sensitive ones for ICS)

Useful webtools: spillover, brightness, ...

BD Spectrum Viewer

The BD Spectrum Viewer is a tool that depicts the excitation and emission curves of fluorochromes common to flow cytometry. This tool can be used to determine appropriate filters to detect a fluorochrome as well as fluorochrome compatibility and fluorescent spillover. Unlike a static image showing excitation and emission histograms, this tool will demonstrate how a fluorochrome will have the same emission profile—but have varying brightness—based on the excitation wavelength of the laser. Choosing different laser wavelengths will transform the emission curve based on the percent excitation at that wavelength.



Useful webtools: spillover, brightness, ...

Flow Cytometry Tools

We offer over a dozen web tools to educate researchers on fluorophores, build the ideal multicolor panel, and choose the right reagents for flow cytometry applications. For additional inquiries, [contact our Technical Services team](#).

Spectra Analyzers

Panel Design Guidance

Fluorophore Information

Pre-Experiment Planning

Post-Experiment Help

Products

Conventional Spectra Analyzer

"Conventional/traditional" flow cytometers use a series of mirrors and filters in order to capture the emission wavelengths from excited fluorophores. Fluors that have emission profiles that fall into the same filter cannot be separated or identified from one another.

Our spectra analyzer can display the excitation and emission spectra for dozens of fluorophores. This allows you to compare the excitation and emission profiles of fluors, custom filters, and laser lines to see what best fits your cytometer.

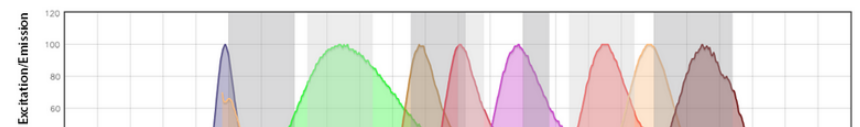
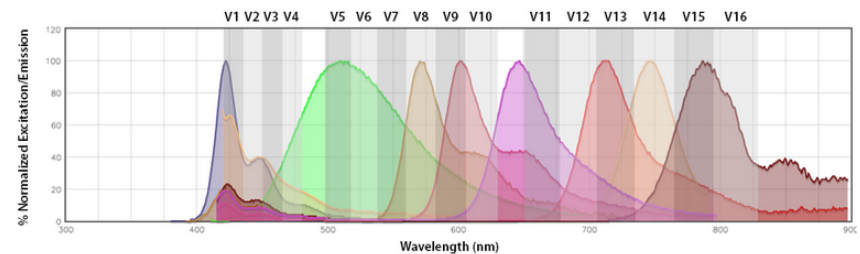
[View the Conventional Spectra Analyzer >](#)

Spectral Cytometry Analyzer

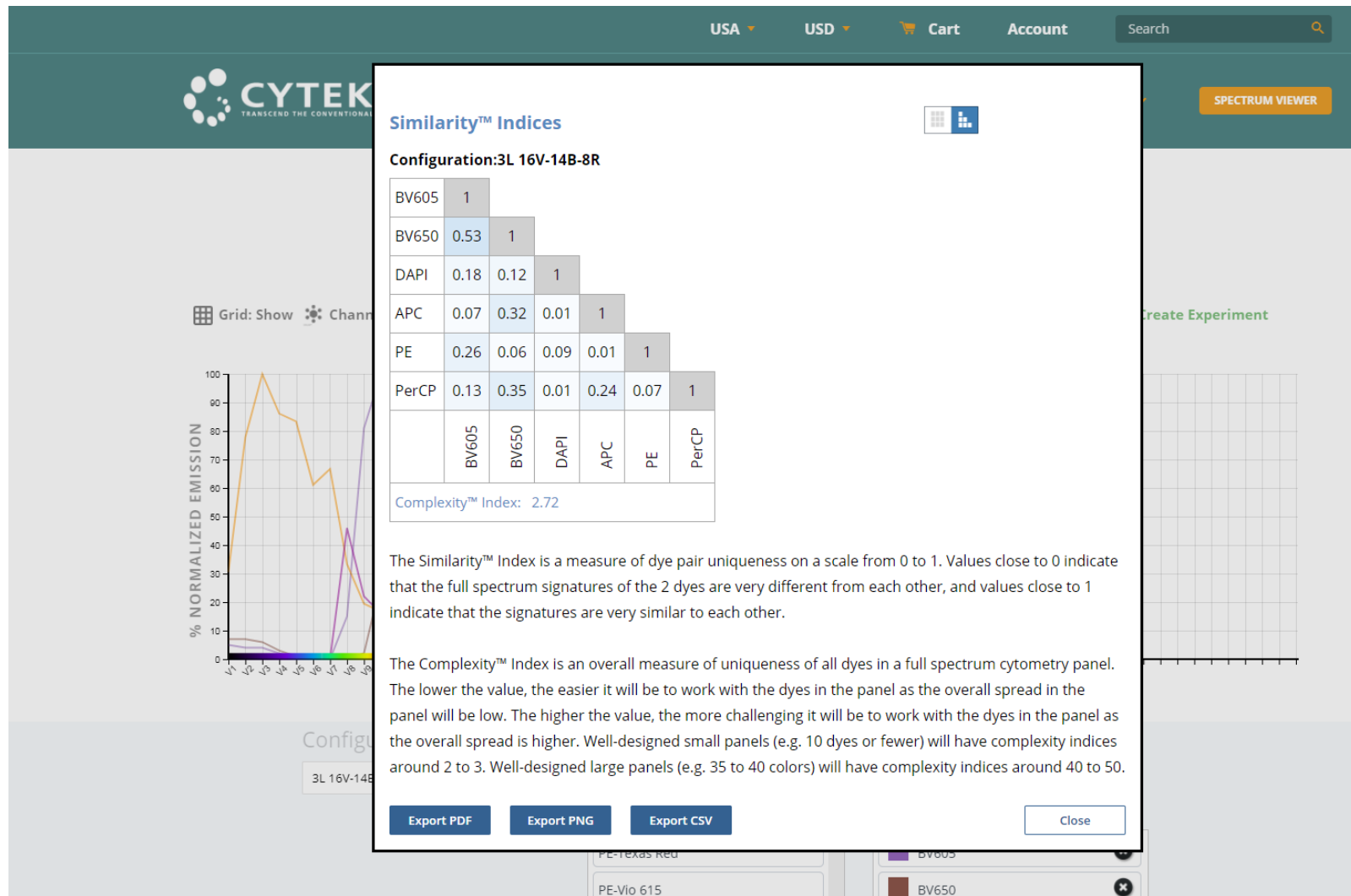
Spectral cytometers capture the full spectrophotometric profile of the fluorophores across all lasers. The fluors' profiles are captured in 10-30 nm segments across the emission range to accurately unmix the fluorophores. This allows fluors that normally cannot be unmixed on a conventional cytometer (e.g. Pacific Blue™ and Brilliant Violet 421™) to be used together on an instrument like Cytek's Aurora.

As such, this web tool is dedicated to displaying the emission spectra of fluorophores based on Cytek's Aurora spectral cytometers.

[View the Spectral Cytometry Analyzer >](#)



Useful webtools: spillover, brightness, ...



Exercise – Identification of human Tregs

1. Which type of cells you want to look at?
 - human Tregs in PBMCs
 - CD3, CD4, CD25, FoxP3
 2. Search for antibodies for tertiary antigens (intranuclear, cytoplasmatic proteins)
 - FoxP3
 - CD25
- Fill your panel

example panel - Tregs

Panel Template BD FACS Aria IIIu					
Laser	Laser Excitation	Parameter	Detector	Filter	Marker:Dye
1	Rot (633 nm)	APC-Cy7, APC-H7, APC-Vio770, APC/Fire750, APC-eFluor780	A	780/60, 735 LP	
		AF700, APC-R700, ZombieNIR	B	730/45, 690 LP	
		APC, AF647, Cy5, eFluor660	C	660/20	
2	Gelbgrün (561 nm)	PE-Cy7, PE-Vio770	A	780/60, 735 LP	
		PE-Cy5, 7-AAD, mPlum	B	670/14, 630 LP	
		PE-TexasRed, PI, mCherry, PE-Dazzle594, ZombieRed, AF594	C	610/20, 600 LP	
		PE, DsRed, Cy3	D	582/15	
3	Blau (488 nm)	PerCP-Cy5.5, PerCP, BB700, PerCP/Vio700, PerCP-eFluor710	A	695/40, 655 LP	
		FITC, GFP/YFP, AlexaFluor488, Zombie Green, VioBright515/FITC, KIRAVIABlue520, AF532	B	530/30, 502 LP	
		SSC	C	488/10	
4	Violet (405 nm)	BV786, BV750, SuperBright780	A	780/60, 750 LP	
		BV650, Qdot655, SuperBright645	B	660/20, 630 LP	
		BV605, Qdot605, SuperBright600	C	616/23, 595 LP	
		BV510, V500, AmCyan, Aqua L/D, eFluor 506, VioGreen	D	530/30, 502 LP	
		DAPI, BV450, BV421, Hoechst, VioBlue, eFluor450, AF430, ZombieViolet	E	450/40	

Exercise – Tregs

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		AF700, APC-R700, ZombieNIR	B	730/45, 690 LP	
		APC, AF647, Cy5, eFluor660	C	660/20	CD25 - APC
2	Gelbgrün (561 nm)	PE-Cy7, PE-Vio770	A	780/60, 735 LP	
		PE-Cy5, 7-AAD, mPlum	B	670/14, 630 LP	
		PE-TexasRed, PI, mCherry, PE-Dazzle594, ZombieRed, AF594	C	610/20, 600 LP	
		PE, DsRed, Cy3	D	582/15	FoxP3 - PE
3	Blau (488 nm)	PerCP-Cy5.5, PerCP, BB700, PerCP/Vio700, PerCP-eFluor710	A	695/40, 655 LP	
		FITC, GFP/YFP, AlexaFluor488, Zombie Green, VioBright515/FITC, KIRAVIABlue520, AF532	B	530/30, 502 LP	
		SSC	C	488/10	
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		BV605, Qdot605, SuperBright600	C	616/23, 595 LP	
		BV510, V500, AmCyan, Aqua L/D, eFluor 506, VioGreen	D	530/30, 502 LP	
		DAPI, BV450, BV421, Hoechst, VioBlue, eFluor450, AF430, ZombieViolet	E	450/40	

Exercise – Tregs

3. Combine and add highly and moderate expressed markers with dim dyes
 - CD3
 - CD4
4. Include live-dead stain
 - fixable

Exercise – Tregs

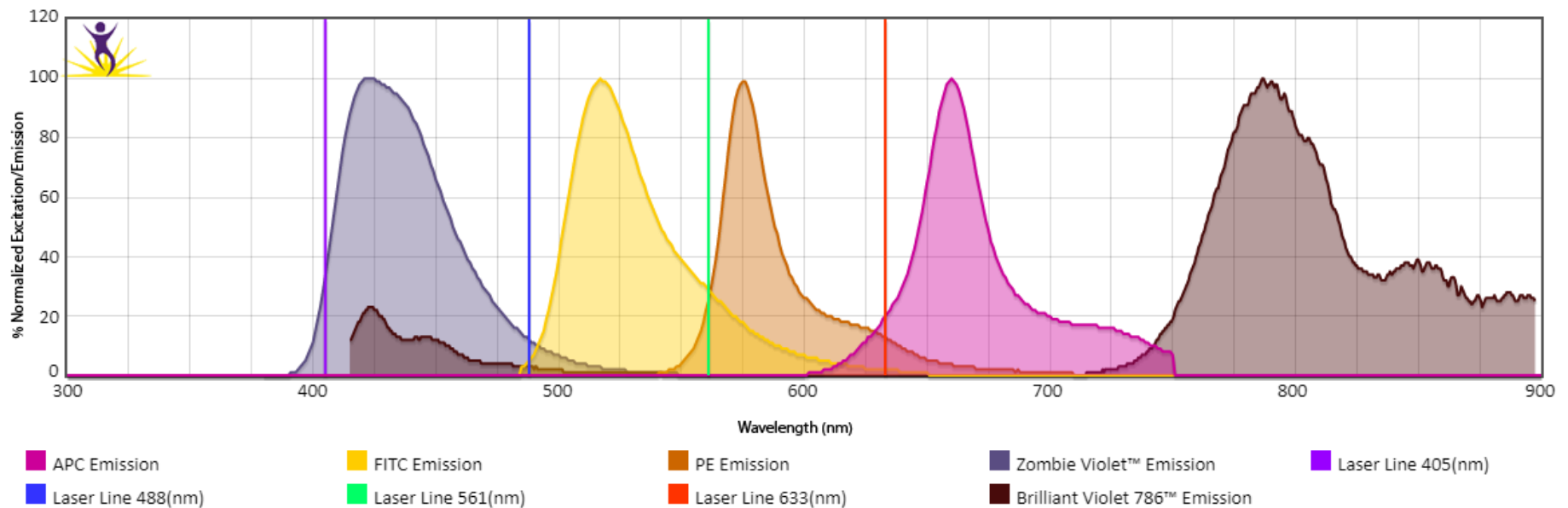
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		APC, AF647, Cy5, eFluor660	C	660/20	CD25 - APC
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		PE-Cy5, 7-AAD, mPlum	B	670/14, 630 LP	
		PE-TexasRed, PI, mCherry, PE-Dazzle594, ZombieRed, AF594	C	610/20, 600 LP	
		PE, DsRed, Cy3	D	582/15	FoxP3 - PE
3	Blau (488 nm)	PerCP-Cy5.5, PerCP, BB700, PerCP/Vio700, PerCP-eFluor710	A	695/40, 655 LP	
		FITC, GFP/YFP, AlexaFluor488, Zombie Green, VioBright515/FITC, KIRAVIABlue520, AF532	B	530/30, 502 LP	
		SSC	C	488/10	
4	Violet (405 nm)	BV786, BV750, SuperBright780	A	780/60, 750 LP	
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Exercise – Tregs

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		FITC, GFP/YFP, AlexaFluor488, Zombie Green, VioBright515/FITC, KIRAVIABlue520, AF532	B	530/30, 502 LP	CD3 - FITC
		SSC	C	488/10	
4	Violet (405 nm)	BV786, BV750, SuperBright780	A	780/60, 750 LP	CD4 – BV786
		BV650, Qdot655, SuperBright645	B	660/20, 630 LP	
		BV605, Qdot605, SuperBright600	C	616/23, 595 LP	
		BV510, V500, AmCyan, Aqua L/D, eFluor 506, VioGreen	D	530/30, 502 LP	
		DAPI, BV450, BV421, Hoechst, VioBlue, eFluor450, AF430, ZombieViolet	E	450/40	Zombie Violet

Exercise – Tregs

5. Check your panel:
- put markers on highly autofluorescent cells in red laser module
 - live-dead stain and ubiquitous markers at the edges of the spectrum (less compensation)
 - compare antibody clones (resolution)
 - use better (tandem-) dyes with less spread (e.g. PerCP → PerCP-Cy5.5 or BB700 or PerCP-Vio700)
 - check for fixability of dyes (use sensitive ones for ICS)



TAKE Home Message

- primary antigens → dim dyes
- secondary antigens → moderate to bright dyes
- tertiary (unknown) antigens → bright to very bright dyes
- spillover spread and cross laser excitation reduces resolution → exchange tandems
- use vendor information, published panels and web tools

Always ask for help or assistance with your panel!!!

Thank you for your attention.

See you next month: **5th September**

Next topic
Cell sorting – why and how?