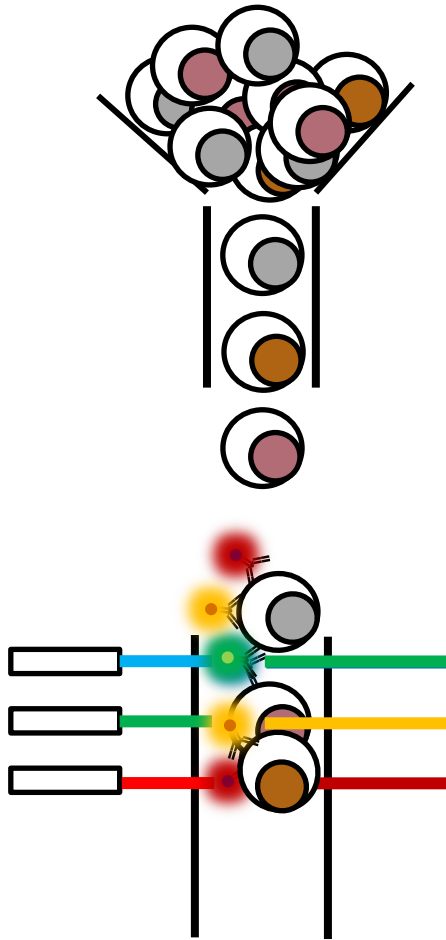


## **5.) „The alpha and omega of preparing cells“**

## **5.) Das A und O der Probenpräparation: Von Asservierung bis Zielzelle**

# Flow cytometry needs single cells



Webinar: Sample Preparation for Flow Cytometry

**Workflow of flow cytometry experiments**

Miltenyi Biotec

Sample preparation      Sample labeling      Acquisition      Analysis

The screenshot shows a webinar slide titled 'Workflow of flow cytometry experiments' by Miltenyi Biotec. The workflow is divided into four stages: 1. Sample preparation, which includes images of a pipette, a piece of tissue, and a multi-well plate. 2. Sample labeling, shown as a test tube containing cells with colored dots. 3. Acquisition, represented by a green flow cytometer. 4. Analysis, shown as a flow cytometry plot with two distinct cell populations. The slide is part of a video player with a progress bar at the bottom showing 1:53 / 39:48.

<https://www.youtube.com/watch?v=mpOIRpIZhPg>

# sample preparation

Garbage in...



...garbage out

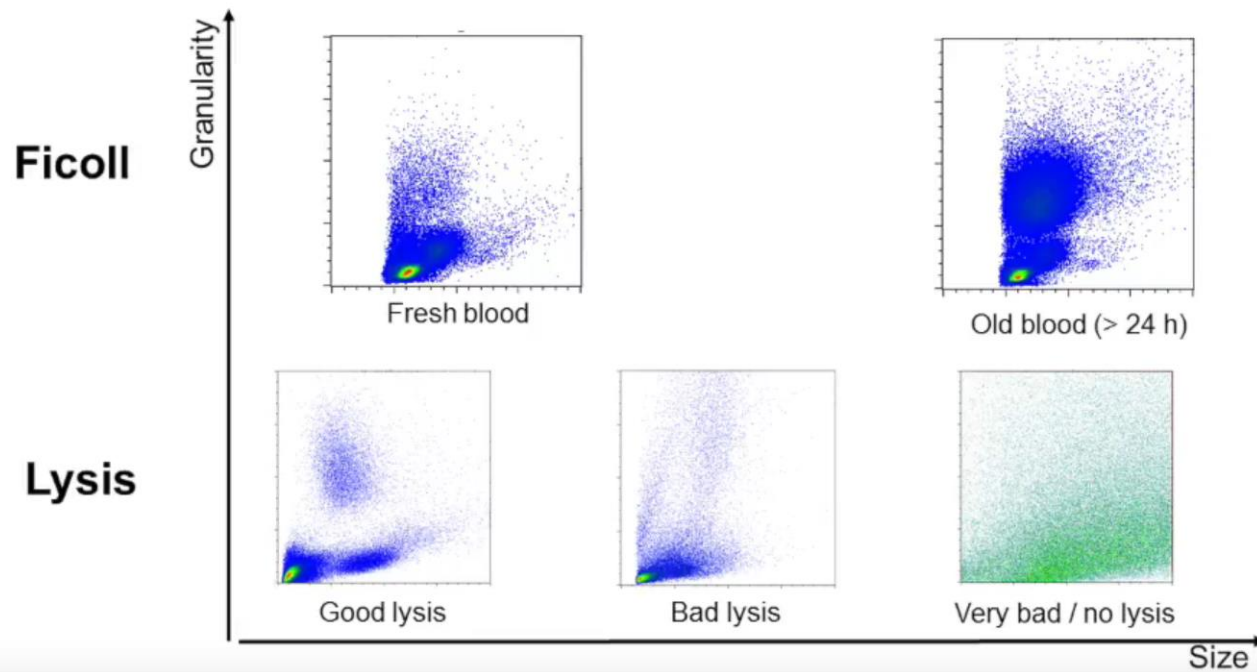
- better sample preparation leads to better results and easy data analysis
- standard procedure:
  - trypsinize cells or lyse blood
  - wash with cell stain buffer (1x PBS 1-2% BSA, 2mM EDTA, 2mM  $\text{NaN}_3$ )
  - stain and measure
- changing BSA and/or EDTA conc. might reduce doublets

# sample preparation – lysing blood

Webinar: Sample Preparation for Flow Cytometry

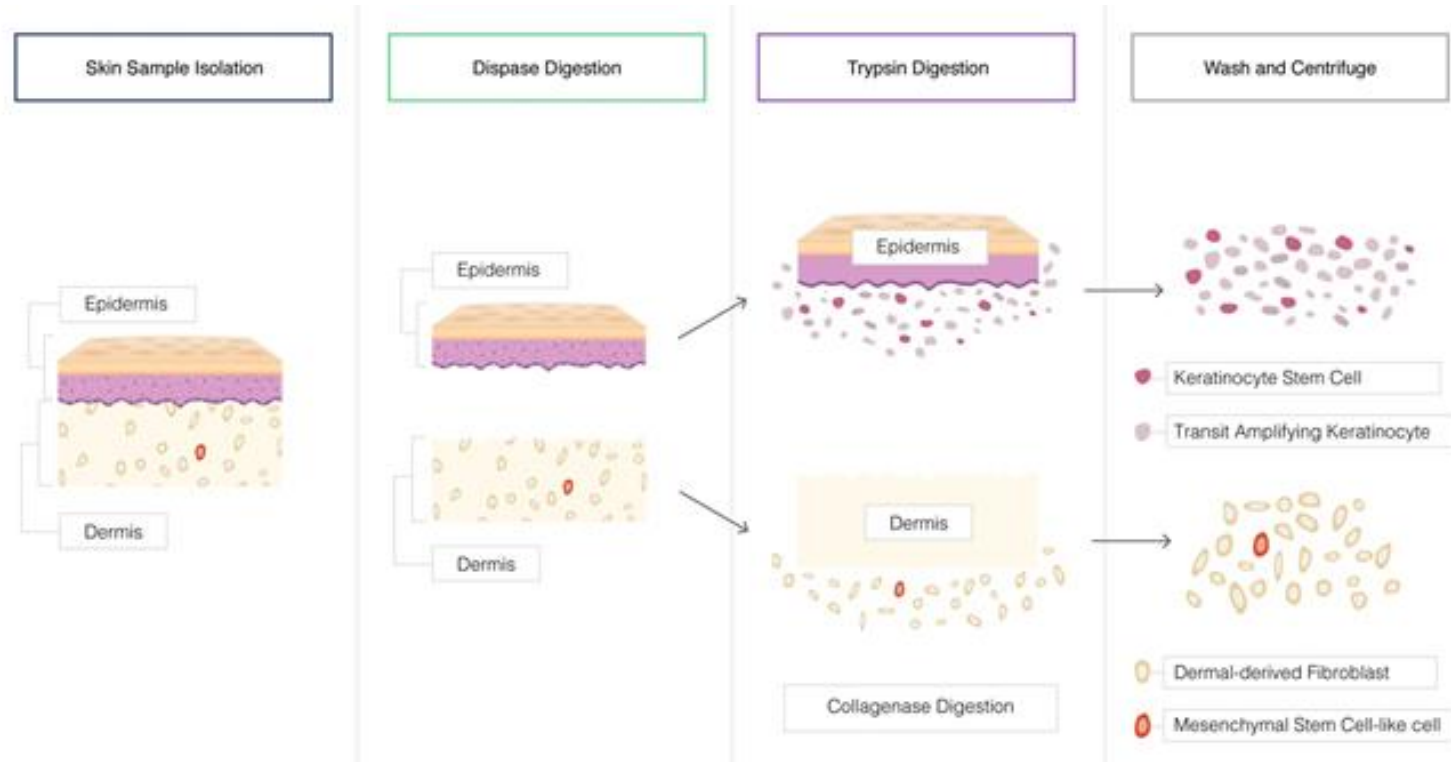


## Preparation of blood



<https://www.youtube.com/watch?v=mpOIRpIZhPg>

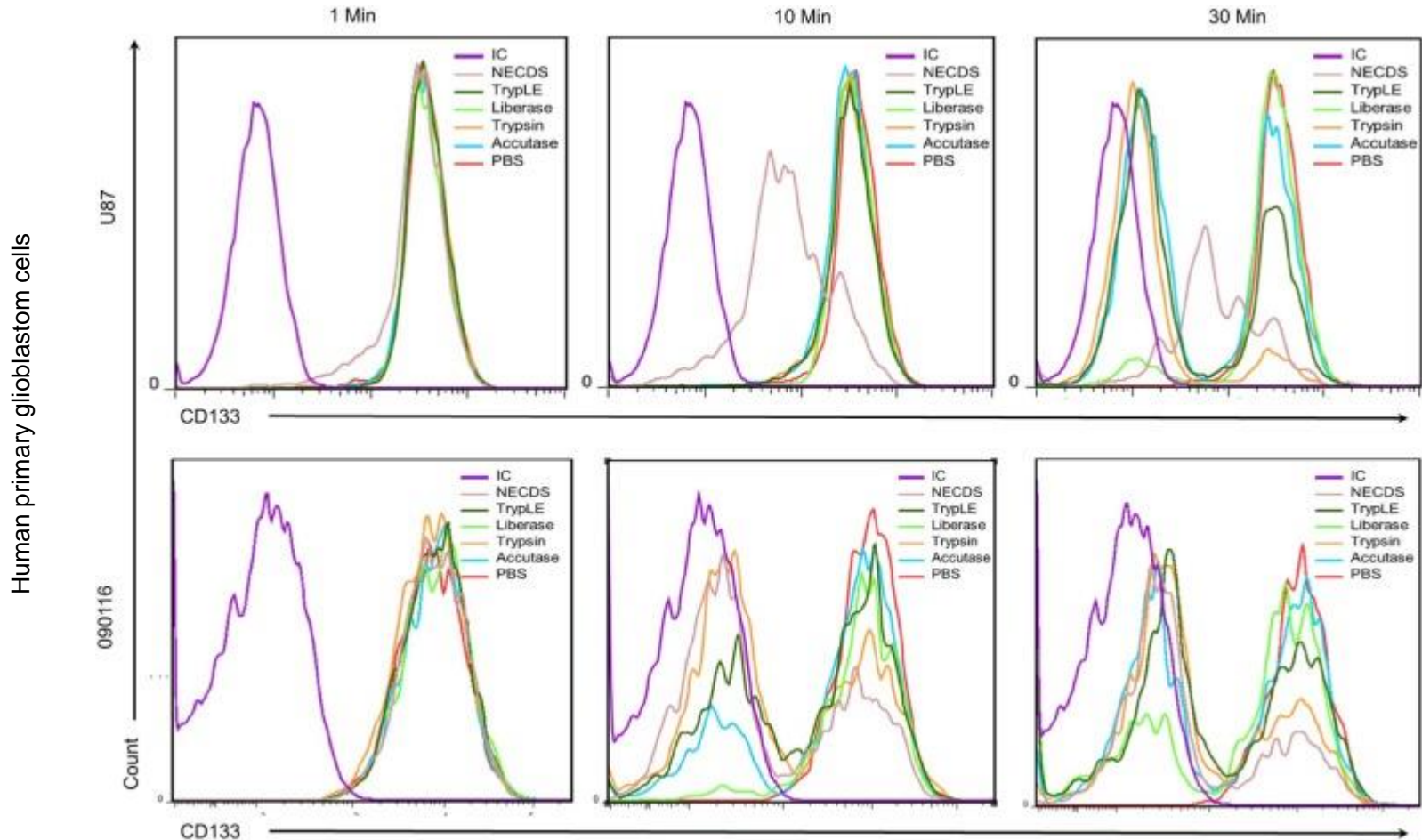
# sample preparation - tissues



<https://www.flowmetric.com/cytometry-blog/sample-preparation-for-flow-cytometry>

- complex enzymatic digestion needed
- goal: single cells with >50% vitality and intact epitopes

# sample preparation - enzymatic digestion



NECDS (non-enzymatic cell dissociation solution): detachment of primary cells, EDTA based

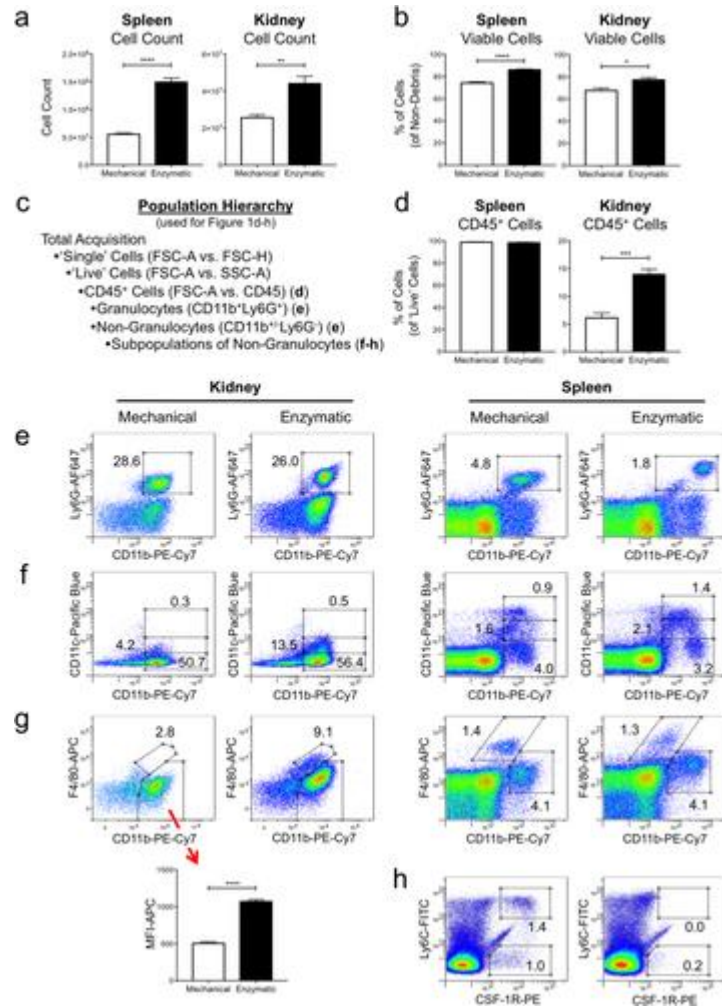
TrypLE (Gibco): animal origin-free, recombinant enzymes for dissociating adherent mammalian cells → replaces trypsin, superior performances

Liberase (Merck): Kollagenase I / II + Thermolysin (Protease)

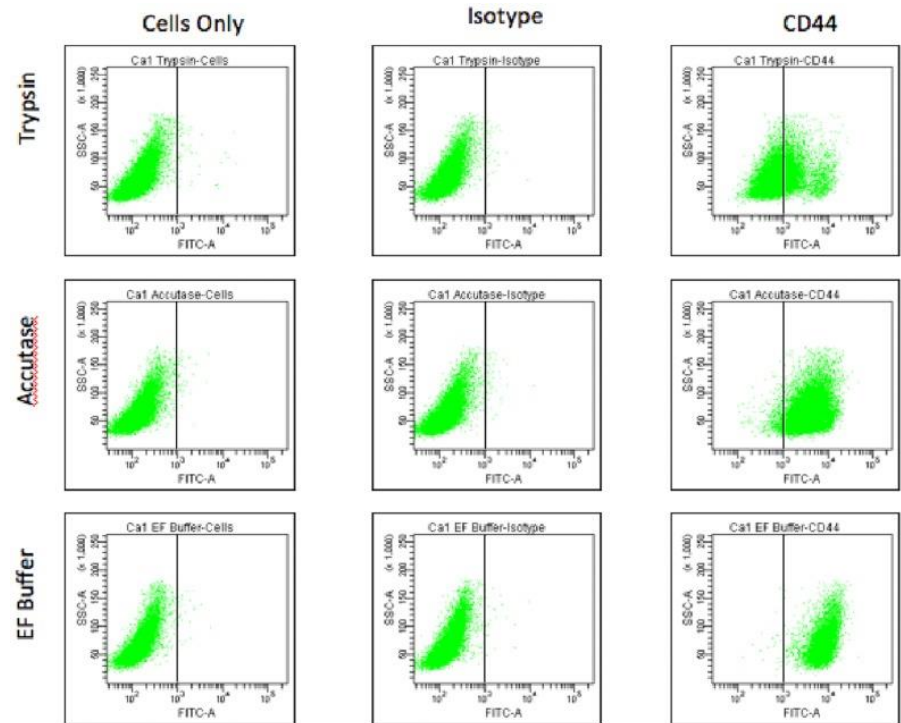
Accutase: animal origin-free, proteolytisch und kollagenolytisch

CD133 – prominin-1 HSC,CSC marker

# enzymatic vs. mechanical dissoziation



- Single cell suspension
- Method of harvest: trypsin, accutase, EDTA  
→ crucial for successful staining



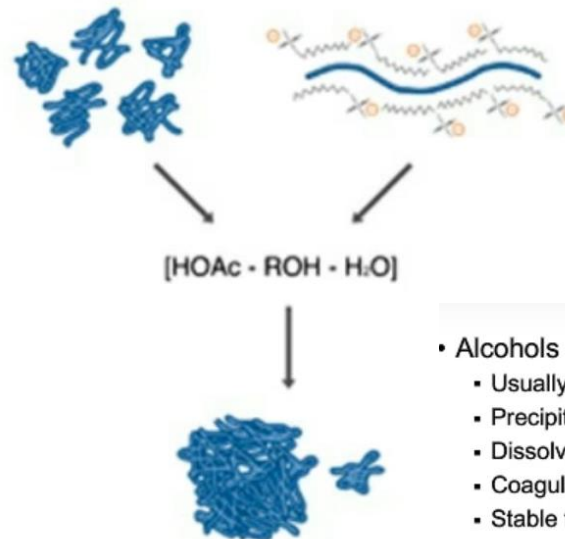
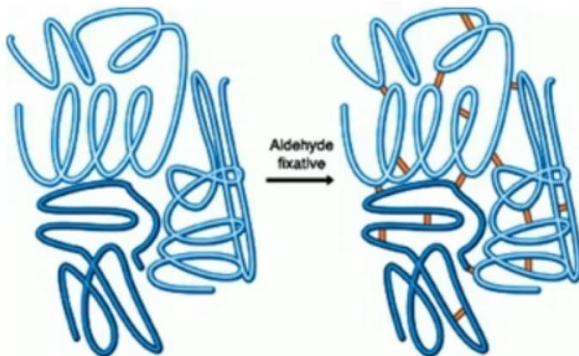
# fixation of cells

- fixation BEFORE staining - epitope alteration
- GFP, PerCP, fluorescence loss after fixation (APC-Cy7 < APC-H7)
- fixation AFTER staining - fluorophore stability
- but pSTAT needs methanol demasking

## • Fixation

### • Formaldehyde

- Create bonds between lysine residues to cross link proteins
- Used between 0.5% and 4%
- Stable for 1-2 weeks but should be kept in PBS after initial fixation



<https://www.youtube.com/watch?v=oeqiCa3nEwc>

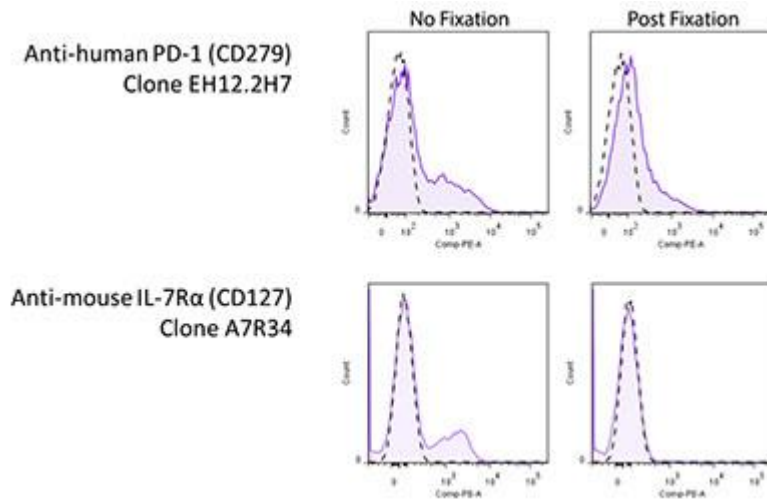


# what to consider - surface staining

- coagulant e.g. EDTA, Heparin interfere with some epitopes
- check enzyme activity and whether targets are „nibbled off“
- Ficoll and RBC Lysis buffer also interfere with surface epitopes
- blocking of Fc receptors when working with PBMCs or any other cell types who express FcR
  - exception: Fc-staining on monocytes, Ig-staining on B cells
- in case of fixation: check tandem fixability
- stimulation reduces or increases expression? →PMA/Iono causes internalization

# what to consider - intracellular staining

- also block intracellularly
- dilute antibodies in perm buffer when perm buffer is saponin (o.a.) based (reversible)
- use bright fluorophors (background/autofluorescence increases always)
- use (FMO) controls



<https://www.biolegend.com/en-us/blog/fix-now-fix-later-considerations-for-the-use-of-paraformaldehyde-fixation-in-flow-cytometry>

Unbound antibodies are trapped in the cells in the case of intracellular staining

The surface antigen is getting internalized

- Wash the cells adequately after each antibody incubation step and include Tween or Triton X in wash buffers.

- Perform all protocol steps at 4°C and use ice cold reagents. We recommend you permeabilize on ice.

# TAKE Home Message

- the better the preparation and protocol of your samples
- the better results/analysis you get
- coagulants, Ficoll, lysis buffer, enzymes, fixation buffer interfere with epitopes and reduce signal
- in multi-colour panels optimization towards high signal, low doublets, high viability is the goal!

Thank you for your attention.

Next topic (6th July): „The golden rules of panel design“

Nächstes Mal (am 6. Juli): „Die goldenen Regeln des Paneldesigns“



## Enzymes for tissue dissociation

Enzyme	Type	Source	Target	
Trypsin	serine protease	bovine/porcine pancreas	predominantly at c-terminal side of AA Lys or Arg	relatively harsh
Papain	cysteine protease	<i>Carica papaya</i>	cleaves basic AAs	relatively harsh, but less digestive than Trypsin
Collagenase	protease, includes also phospholipases or neuraminidases	<i>Clostridium histolyticum</i>	peptide bonds in collagen; Different types available, depending on supplier Roche: A, B, D, H, P Sigma-Aldrich: I-IV, Worthington: Type 1-4	different collagenase types with different levels of unspecific activities
Liberase Blendzymes	blend of purified collagenases with neutral proteases	<i>Clostridium histolyticum</i> and <i>Bacillus polymyxa</i> or <i>B. thermoproteolyticus</i>	Supplied by Roche, more defined activity and composition	developed for pancreatic islet transplantation research
Dispase	protease	<i>Bacillus polymyxa</i>	fibronectin, collagen IV (collagen I)	separation of dermis and epidermis
DNase	nuclease DNase I: neutral DNase II; functions optimally at acidic pH	bovine pancreas	DNA	avoid cell clumping

<https://www.youtube.com/watch?v=mpOIRpIZhPg>