

**Cell cycle analysis – simple but  
complicated?**

**Zellzyklusanalysen – einfach und doch  
kompliziert?**

# Cell cycle analysis – simple but complicated?

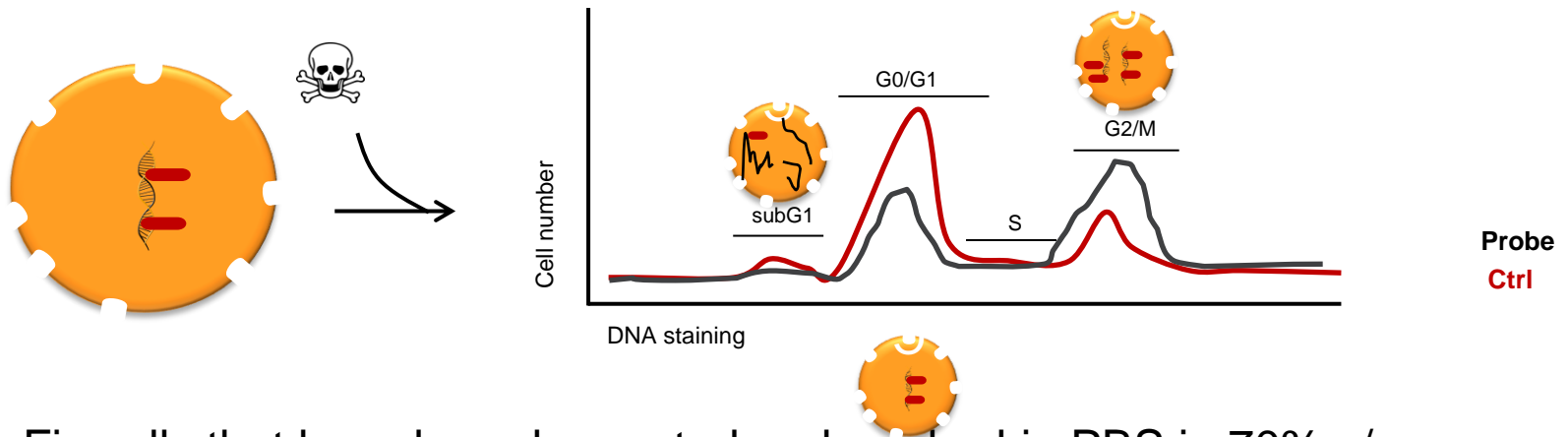
simple:

- use of intercalating DNA-Dyes
- only one colour needed → can be done on nearly any flow cytometer
- one of first applications of flow cytometry in history

complicated:

- many different protocols available
- small differences in treatment can have big effects
- RNA, doublets and clumps disturb cell cycle analysis

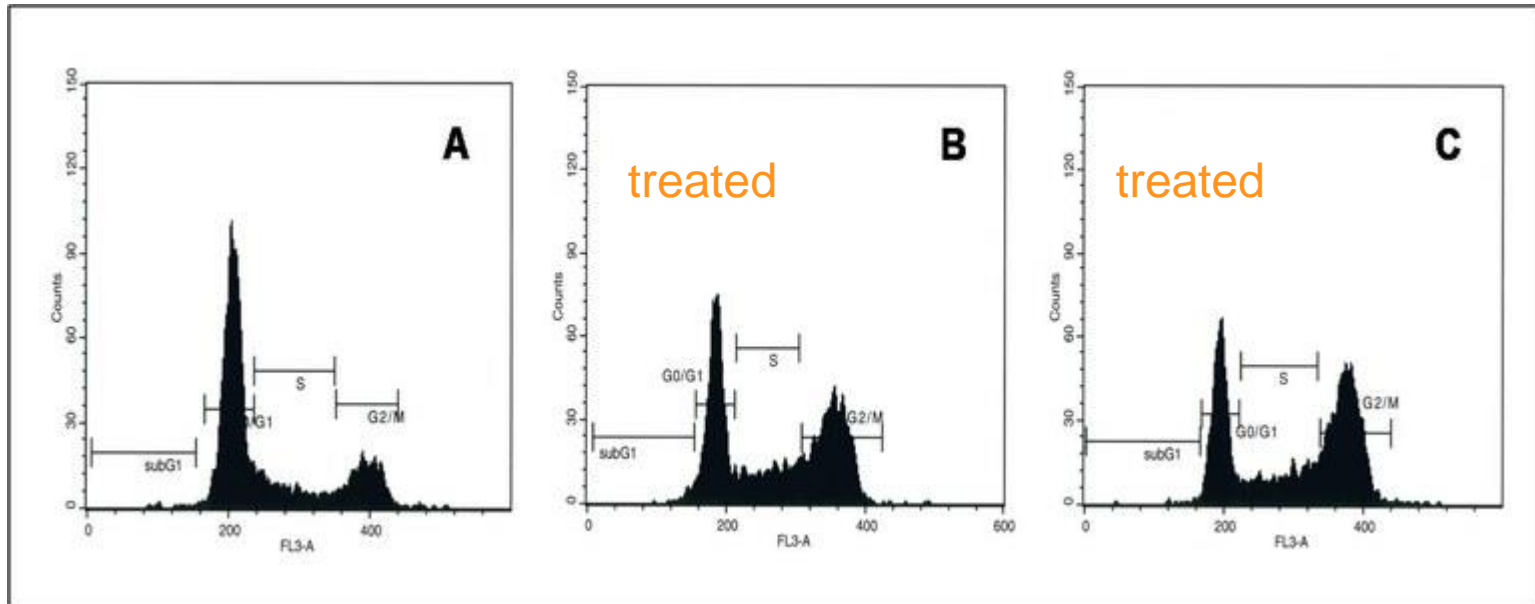
# principle and general steps



1. Fix cells that have been harvested and washed in PBS in 70% v/v ethanol. Adding the ethanol dropwise to the cell pellet while vortexing will ensure that all cells are fixed and will minimize clumping.
2. Fix cells for 30 min at 4°C, after which wash cells twice in PBS (850 × g). Be careful to avoid cell loss when discarding the supernatants.
3. Treat cells with RNase (50 µL, 100 µg/mL) in order to ensure that only DNA is stained.
4. Add PI (200 µL PI, 50 µg/mL stock solution) immediately before analyzing.

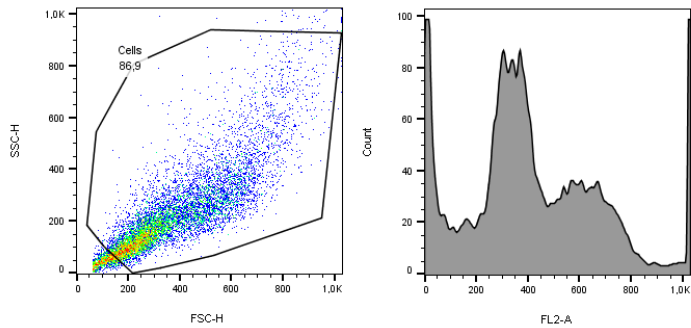
Cossarizza, A et al. "Guidelines for the use of flow cytometry and cell sorting in immunological studies" (second edition). (2019), Eur. J. Immunol., 49: 1457-1973. <https://doi.org/10.1002/eji.201970107>

# Example of cell cycle arrest

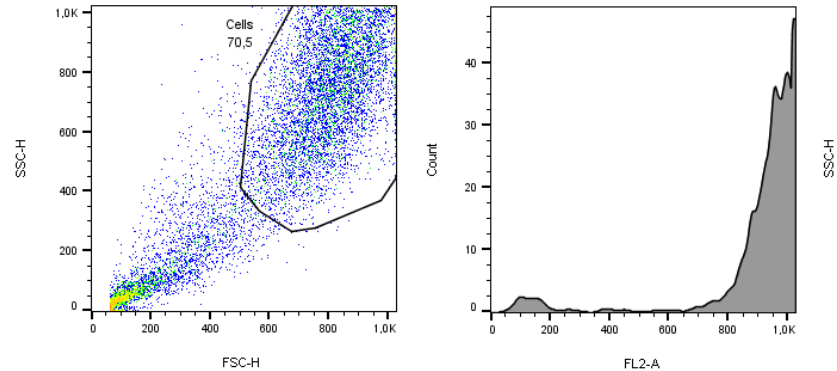


<https://www.uni-giessen.de/fbz/fsp/meu/methodenplattform/analysen2/zellzyklusanalysen>

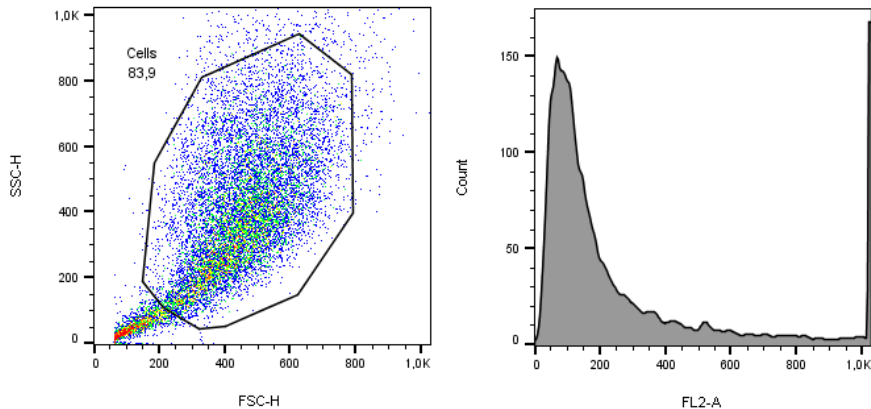
# Cell cycle analysis – What had happened here?



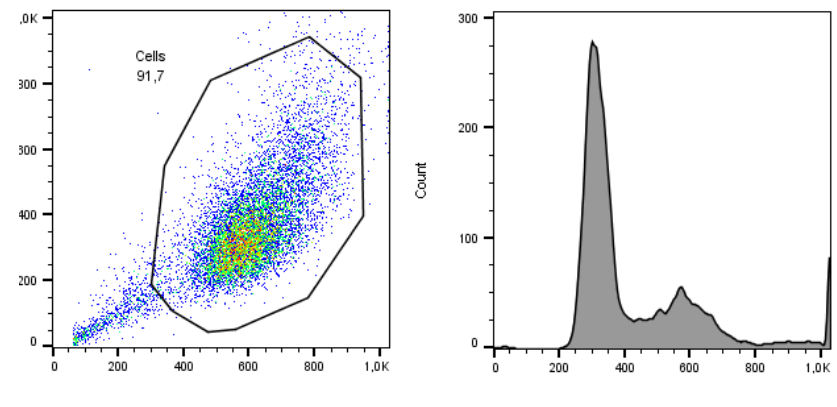
too many dead cells,  
low number of living cell



PMTV for scatter and dye not set properly



too much PI added



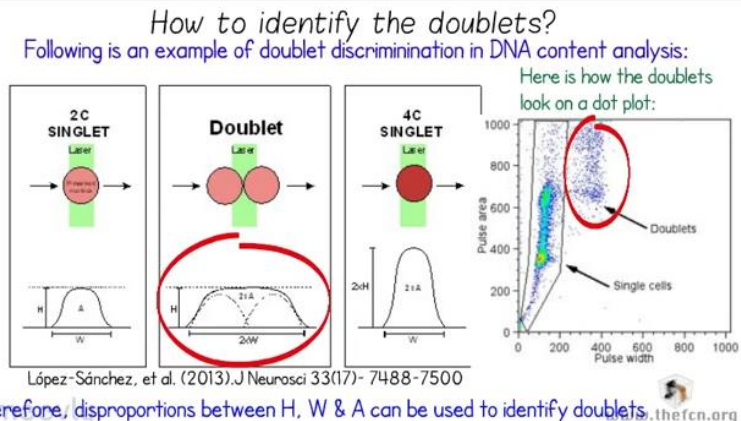
no doublet exclusion

# What to consider?

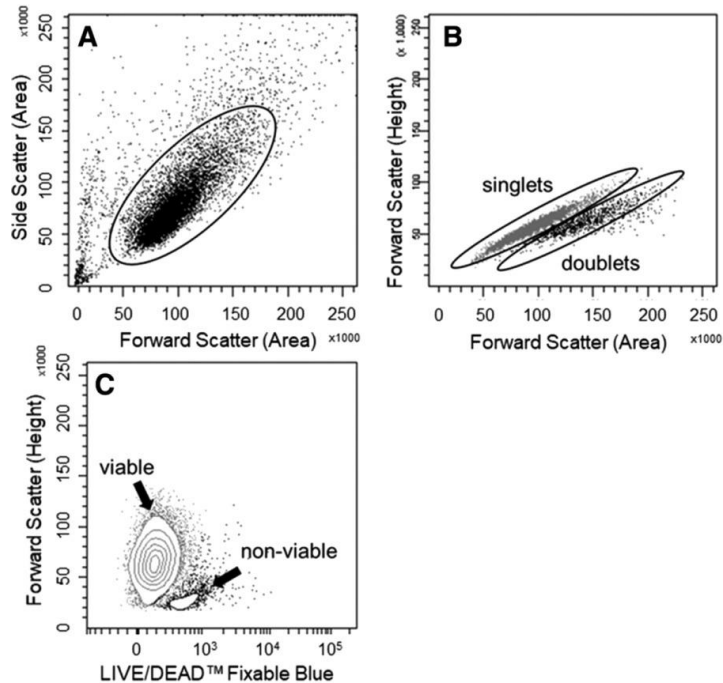
- Many DNA Dyes available:
  - PI, Hoechst stains, TO-PRO-3, SYTOX, acridine orange, pyronin Y, 7-AAD, DAPI, DRAQ5™, and DRAQ7™
  - bind RNA (PI), others less (7-AAD, DAPI), staining of living cells (DRAQ Dyes)
- PFA-fixation reduces resolution, because of chromatin crosslinking
- Always count cells before DNA staining →  
DNA-dye – cell – ratio should be stable

# What to consider?

- exclude doublets
- if possible exclude dead cells  
(scatter or better fixable live-dead-stain)
- lower flow rate results in more accurate acquisition → sharp peaks
- measure & analyze with linear scaling
- record 10.000 – 20.000 cells
- adjust G1 peak to a definite value, e.g. ~ 200, and check for the G2/M to appear at the doubled value, e.g. 400 (doubling of DNA content)



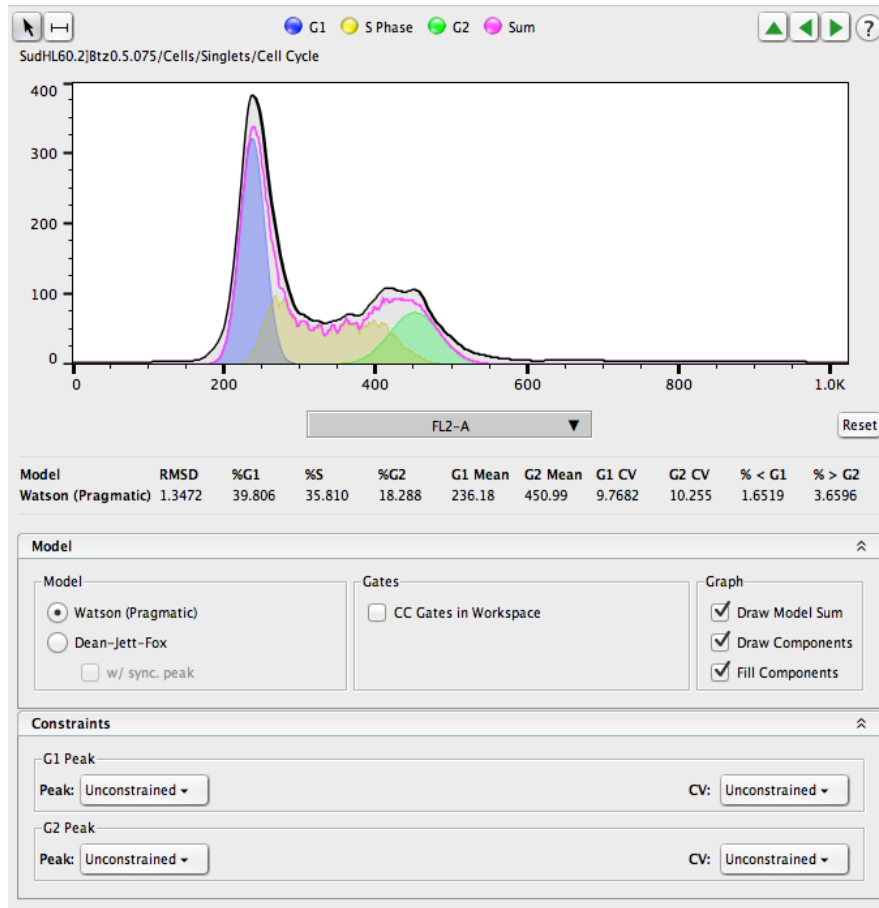
# experimental improvements



- adjust concentration of treatment  
(too many apoptotic cells)
- include live-dead-staining
- include untreated control (always same confluency)
- maybe starve and synchronize cells before assays start



# cell cycle analysis tools



- Specialized programs such as ModFit LT™ from Verity Software House (<http://www.vsh.com/products/mflt/mfFeatures.asp>) and Multicycle AV™ from Phoenix Flow Systems (<http://www.phnxflow.com/MultiCycle.standalone.html>) have been designed for this purpose.

European Journal of Immunology, Volume: 49, Issue: 10, Pages: 1457-1973, First published: 21 October 2019, DOI: (10.1002/eji.201970107)

<https://docs.flowjo.com/flowjo/experiment-based-platforms/cell-cycle-univariate/>

# cell cycle analysis –TAKE HOME MESSAGE

- improve data quality:
  - dye selection and protocol optimization
  - doublet exclusion and low flow rate
  - because of linear scale small differences have big effect
  - stable cell number and dye concentration needed
- FlowJo provides easy analysis tool
  - needs clear G1-peak
  - good untreated control necessary
- ‚easy‘ cell cycle analysis can be more complicated than thought

**Thank you for your attention.**

**See you next month 04<sup>th</sup> May.**

**Next topic:  
Aggravating compensation –  
discover errors and correct them**

**Die leidige Kompensation – Fehler erkennen und  
nachbessern**